**foodproof®**

**Celery Detection Kit**

**Quick Reference Procedure**

- **5’Nuclease** -

Order no. R 302 60  

Version 2, June 2017

PCR kit for the qualitative detection of celery DNA using real-time PCR instruments. Before starting, it is strongly recommended to read the entire product manual available on our website.

**PROGRAM SETUP**

Program your real-time PCR instrument before setting up the PCR reactions. Select the following channels:

- FAM (celery) and HEX (Internal Control).

![PCR Reaction Program](image)

**Pre-incubation:** 1 cycle
- **Step 1:** 37 °C for 4 min
- **Step 2:** 95 °C for 10 min

**Amplification:** 50 cycles
- **Step 1:** 95 °C for 5 sec
- **Step 2:** 60 °C for 60 sec

* Fluorescence detection

For some real-time PCR instruments the probe quencher as well as the usage of a passive reference dye has to be specified. This kit contains probes with a non-fluorescent “dark” quencher and no passive reference dye.

**DATA INTERPRETATION**

Verify results of positive (Control Template) and negative controls (H₂O), before interpreting sample results. Always compare samples to positive and negative control. Review data from each channel and interpret results as described in the table.

<table>
<thead>
<tr>
<th>FAM</th>
<th>HEX</th>
<th>Result Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>+</td>
<td>+ or -</td>
<td>Positive for celery</td>
</tr>
<tr>
<td>-</td>
<td>+</td>
<td>Negative for celery</td>
</tr>
<tr>
<td>-</td>
<td>-</td>
<td>Invalid</td>
</tr>
</tbody>
</table>

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PREPARATION OF THE PCR MIX

Take appropriate precautions to prevent contamination, e.g. by using filter tips and wearing gloves. Thaw reagents, mix (do not vortex!), and briefly spin vials before opening.

1. ADD PCR MIX
Pipet 20 µl of Master Mix into each strip or plate well (n samples + 2 controls).

2. ADD SAMPLES AND CONTROLS
Pipet 5 µl of samples, negative control (colorless cap) or Control Template (purple cap) into respective wells.

OPTIONAL: STANDARD CURVE
For quantification prepare standard curve. Please refer to Allergen RM 800 (Order No. A 500 13) product manual.

3. SEAL
Seal strips/plate accurately.

4. CENTRIFUGE
Briefly spin strips/plate in a suitable centrifuge.

5. START REAL-TIME PCR RUN
Cycle samples as described above.